

Bicyclic Azasugars Containing a Glycosidic Heteroatom: D-Xylose Analogues

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Abstract: Bicyclic azasugar analogues of D-xylose containing a glycosidic heteroatom have been prepared and characterized by X-ray crystallography and NMR spectroscopy. Compounds in which the glycosidic heteroatom is N (substituted with H or CH₃), O, and S have been prepared, and the ring fused to the azasugar ring has been 5- or 6-membered; 7-membered analogues are either unstable or do not form. α -Anomers are present when the glycosidic heteroatom is O or S but not N, and even then they are less favored than the corresponding β -anomers. © 1999 Elsevier Science Ltd. All rights reserved.

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Naturally-occurring compounds which resemble sugars but have a nitrogen atom in place of the ring oxygen of sugars have been termed azasugars. Because many of these compounds inhibit glycosidases, they have been the subject of substantial interest.^{1,2} These azasugars frequently have been the objects of synthetic efforts, and, in addition, a great many analogues of the natural products have also been prepared with the aim of finding compounds with improved inhibitory properties. In the development of glycosidase inhibitors, it is very desirable to discover ways to produce selectivity for a single class of enzymes or even among members within a class. The majority of azasugars reported so far consist of either one ring or two rings which share a nitrogen atom. Most have lacked a heteroatom comparable to the glycosidic oxygen atom present in sugars. It is well known that glycosidases are exquisitely selective regarding the anomeric configuration of their substrates and are consequently named based on that selectivity, for example, as α -glucosidases. selectivity is apparently a consequence of the placement within the active sites of these enzymes of catalytic carboxylic acid residues which serve to protonate the glycosidic oxygen atom. A few azasugars have been reported which contain a singly-bonded glycosidic heteroatom; these include the natural products nojirimycin 1³ and related compounds mannojirimycin⁴ and galactostatin⁵ (all three exist as mixtures of anomers), the unusual natural product kifunensine 2, ⁶ synthetic 1-β-amino-1-deoxynojirimycin 3, ⁷ and thia-analogues such as 48 and 5.9 More recently we have reported the preparation of bicyclic diazasugars such as 6.10.11 It appears that those azasugars with a singly-bonded glycosidic heteroatom and a strong preference for a single anomeric configuration (2, 3, and 6) are very selective inhibitors of the glycosidases that cleave glycosides of the same anomeric configuration; those azasugars that are mixtures of anomers (1 and related compounds) or have no anomeric heteroatom (1-deoxy analogues) are not selective and inhibit both α - and β -glycosidases. Azasugars which possess an anomeric heteroatom may act as pseudosubstrates of glycosidases, and this may account for their inhibitory selectivity.

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In light of these observations, we have set as research objectives the preparation of bicyclic azasugars with nearly exclusively α -configurations and others which are essentially only β -anomers. Our previous experience with octahydropyrido[1,2-a]pyrimidines such as 6 has shown that the "glycosidic" nitrogen atom has an extremely strong preference for an equatorial orientation relative to the piperidine ring. For example, 7 and 8 are the preferred conformations for the α -D-ribose and β -L-arabinose analogues, respectively; two axial hydroxyl groups are preferred over one and an axial NHR group which is consistent with reported A-values for these groups¹² and the possible involvement of an exo-anomeric effect. Based on the presumed desirability of mimicking a sugar glycoside in the ${}^{1}C_{4}$ conformation, we have also set conformational control as another of our objectives. Herein we report the syntheses, configurations and conformational preferences of D-xylose azasugar analogues 9 in which X and n have been varied.

Our previous syntheses of bicyclic diazasugars have involved initial preparation of sugar aminals followed by internal displacement of a hydroxyl group which was activated *in situ*. Application of this method to prepare bicyclic azasugars that were not octahydropyrido[1,2-a]pyrimidines failed when attempts to prepare the requisite heterocycles 10 led to the xylosylamines 11 instead. A simple approach to overcoming this problem involved blocking and concomitantly activating the primary hydroxyl group of xylose (e.g. 12) to allow both formation of the requisite heterocycles 13 and promote the final cyclization reactions to give the desired products 9.¹⁴

HO,
$$(CH_2)_n$$
HO $(CH_2)_n$

The synthesis of a terminally blocked and activated D-xylose derivative began with readily available 1,2-O-isopropylidene-5-O-tosyl-α-D-xylofuranose 14. Although 14 itself could be deblocked to 15, a functional precursor to the final products, the instability of 15 prompted the conversion of 14 to bromide 16 followed by deblocking with trifluoroacetic acid to the more stable bromide 17. ¹⁵

To confirm the utility of this synthetic approach, an aqueous solution of 17 was treated with 1,3propanediamine to give known bicyclic diazasugar 6 which could be isolated by simply treating the reaction mixture with an anion-exchange resin to remove HBr, evaporation to dryness, and crystallization. Using this method with 3-amino-1-propanol produced 18, the oxygen analogue of 6, as a mixture of anomers. The βanomer (18β) crystallized from a concentrated solution of the mixture in ethanol. Upon dissolution of the crystals in D₂O, an equilibrium mixture containing 12% of the α -anomer (18 α) was established within less than three days. Analysis of ¹H-¹H three-bond coupling constants for the protons of the azasugar ring (see Table 1) established the conformations of the anomers as shown in 18α and 18β ; due to signal overlap, in some instances a coupling constant for 18\alpha was obtained from only one signal. In contrast to compounds 7 and 8 which prefer conformations with the anomeric nitrogen atom equatorial to the piperidine ring, compound 18α prefers to have the anomeric oxygen atom axial. This must be a consequence of the presence of a strong endoanomeric effect when the anomeric atom is oxygen and the requirement of having three hydroxyl groups be axial in order for the anomeric oxygen atom to be equatorial. In contrast to compound 6 which is only the βanomer, the occurrence of some of the α-anomer of 18 must be due to the endo-anomeric effect compensating for the repulsive steric interactions introduced by the axial anomeric oxygen atom. Both anomers of 18 mimic the preferred conformations of the anomers of D-xylose.

Table 1. Selected ¹H-¹H NMR Coupling Constants of Bicyclo[4.4.0] azasugars in D₂O

Comp	ounds	$J(\mathrm{Hz})$						
No.	X	$J_{ m 6ax,6eq}$	$J_{ m 6eq,7}$	$J_{ m 6ax,7}$	$J_{7,8}$	$J_{8,9}$	$J_{9,9a}$	
6	NH	11.7	4.9	11.0	9.5	9.3	8.9	
18α	0	11.0	4.9	11.0	9.8	9.8	3.1	
18 β	0	11.7	4.9	10.7	9.3	9.4	7.6	
19	NCH ₃	11.7	4.9	10.7	9.3	9.3	8.8	
20		11.7	5.0	9.3	9.3	9.3	9.3	

Since none of the α-anomer of compound 6 was observed, preparation of an N-substituted analogue was undertaken to determine the effect of this substitution on anomeric preference. Compound 17 was treated with N-methyl-1,3-propanediamine, and a mixture of two products in about a 5:1 ratio was obtained from which the major was obtained pure by repeated crystallizations. ¹H NMR spectral data (Table 1) showed this compound to be 19 which was present in solution as only the β-anomer. X-ray structure analysis (see Figure 1 and Table 2) confirmed this structure and showed the methyl group to be axial in the crystalline state. The conformation of 19 in solution was also confirmed by observation of Nuclear Overhauser Effects (NOE's) between H-6ax, H-8, and H-9a, and the orientation of the methyl group in solution was also determined to be primarily axial by the observation of NOE's between it and the H-3ax and H-9 protons. Such a preference for an axial N-methyl group has been previously observed in related heterocyclic systems 16 and may be ascribed to the presence of an exo-anomeric effect; from the X-ray data, the length of the N₁-C_{9a} bond was determined to be slightly shorter (0.013 Å) than that of the N₅-C_{9a} bond which is consistent with an exo-anomeric effect.¹⁷ The minor component formed in this reaction could be favored over 19 in a ratio of 2:1 by the addition of three equivalents of triethylamine to the reaction. It was then possible to purify it by crystallization. 'H NMR spectral data (Table 1) suggested that the minor component was also a β-anomer, and its structure was determined by X-ray crystallography to be 20 (see Figure 2 and Table 2); a very strong exo-anomeric effect is suggested to be present in this compound by the much shorter (0.177 Å) length of the N_1 - C_{9a} bond than that of the N_5 - C_{9a} bond. The influence of triethylamine on the product ratio may be a consequence of it scavenging HBr and freeing the more basic and more nucleophilic methylamino group for participation in the displacement reaction.

17
$$\frac{\text{NH}_2 \text{ NHCH}_3}{\text{H}_2 \text{O}}$$
 $\frac{\text{HO}}{\text{HO}}$ $\frac{\text{NI}}{\text{N}}$ $+$ $\frac{\text{HO}}{\text{HO}}$ $\frac{\text{NI}}{\text{N}}$ $+$ $\frac{\text{HO}}{\text{HO}}$ $\frac{\text{NI}}{\text{N}}$ $\frac{\text{HO}}{\text{N}}$ $\frac{\text{NI}}{\text{N}}$ $\frac{\text{HO}}{\text{N}}$ $\frac{\text{NI}}{\text{N}}$ $\frac{\text{HO}}{\text{N}}$ $\frac{\text{NI}}{\text{N}}$ $\frac{\text{HO}}{\text{N}}$ $\frac{\text{NI}}{\text{N}}$ $\frac{\text{$

To test the effect of ring size on anomeric preference, the preparation of analogues in which n = 0 (see 9) was undertaken. Treatment of 17 with 2-aminoethanethiol gave sulfur analogue 21 as a mixture of anomers

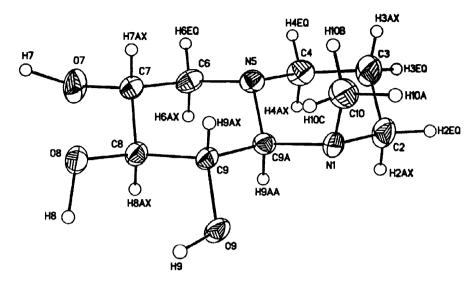


Figure 1. X-ray Structure of $[7R-(7\alpha,8\beta,9\alpha,9a\alpha)]$ -Octahydro-1-methyl-2*H*-pyrido [1,2-*a*]pyrimidine-7,8,9-triol (19)

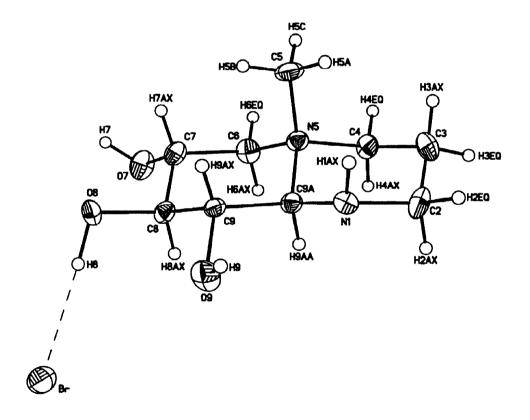


Figure 2. X-ray Structure of $[5S-(5\alpha,7\beta,8\alpha,9\beta,9a\beta)]$ -Octahydro-7,8,9-trihydroxy-5-methyl-2*H*-pyrido[1,2-a]pyrimidin-5-ium bromide (**20**)

with the α -anomer (21 α) constituting 30% of the mixture at equilibrium. As with the oxygen analogue, the β -anomer crystallized. Three-bond ${}^{1}H^{-1}H$ coupling constants (Table 3) for the anomers were consistent with the conformations shown in 21 α and 21 β ; difference NOE's were used to confirm the conformational assignments and distinguish between H-3a and H3b. Like 18 α , the α -anomer prefers to have the glycosidic atom axial.

17
$$\frac{H_2N}{H_2O}$$
 $\frac{HO}{HO}$ $\frac{N}{HO}$ $\frac{HO}{HO}$ $\frac{N}{HO}$ $\frac{N}{S}$ $\frac{21\alpha}{S}$

Table 2. Crystal Data and Summary of X-ray Experimental Conditions^a

Compound	19	20		
Formula	$C_9H_{18}N_2O_3$	C ₉ H ₁₉ BrN ₂ O ₃		
Formula Weight	202.25	283.17		
F (000)	1320	584		
Crystal size, mm	$0.7 \times 0.3 \times 0.08$	$0.5 \times 0.22 \times 0.16$		
μ , mm ⁻¹	0.096	3.577		
Crystal system	Orthorhombic	Orthorhombic		
Space group	P2 ₁ 2 ₁ 2 ₁	$P2_{1}2_{1}2_{1}$		
a, Å	8.804 (2)	6.4679 (6)		
b, Å	11.956 (4)	12.832 (2)		
c, Å	29.854 (11)	13.812 (2)		
V , \mathring{A}^3	3143	1146.3		
Z	12	4		
Dx, g/cc	1.283	1.641		
20 range, deg	4.0 to 45.0	4.0 to 50.0		
Independent data	2536 (Rint = 0.0219)	1231 (Rint = 0.041)		
Data/restraints/parameters	2507/0/380	1231/0/139		
Final R indices [I>2σ (I)]	$R_1 = 0.0528$, $wR^2 = 0.1176$	$R_1 = 0.0339$, $wR^2 = 0.0812$		
Goodness of fit on F ²	1.096	1.051		
Largest peak and	0.258	0.565		
hole in difference map, eÅ ⁻³	-0.244	-0.510		

^a Tables containing the structure determination summary, atomic positional and thermal parameters, bond lengths, and bond angles for these compounds have been deposited in the Cambridge Crystallographic Data Center. These data can be obtained from the Director, Cambridge Crystallographic Center, University Chemical Laboratory, Lensfield Road, Cambridge, CB2 IEW, U.K.

When 17 was treated with 1,2-ethanediamine, two products formed which could not be separated. However, it was possible to assign conformational structure 22 (a β -anomer) to the major component based on analysis of its ^1H - ^1H NMR coupling constants (Table 3). It was not possible to definitively assign the structure of the minor compound, but based on its ^{13}C NMR signals, it appears to be ring-contracted analogue 23 which might arise through the intermediacy of an epoxide. When 17 was treated with 2-aminoethanol, a mixture of products formed that appeared to be primarily the anomers of compound 24. The latter had very limited stability and could not either be purified or, consequently, characterized by ^1H NMR. However, quantitative ^{13}C NMR and ^{13}C chemical shift comparison with 18α and 18β led to the conclusion that 24 was a 1:2 mixture of α - and β -anomers. This was based on two characteristics of the ^{13}C NMR spectra of 18α and 18β and also xylosides and glucosides: the anomeric carbon and the 3-position carbon of the β -anomers appear considerably farther downfield than the corresponding carbons in the α -anomers or any other carbons in these

types of compounds. The major anomer had peaks at 94.3 and 77.2 ppm while the minor had a peak at 92.4 ppm with the next highest at 73.9 ppm.

Table 3. Selected ¹H-¹H NMR Coupling Constants of Bicyclo[4.3.0]azasugars in D₂O

Compounds		J (Hz)						
No.	X	$J_{ m 5ax,5eq}$	$J_{ m 5eq.6}$	$J_{5ax.6}$	$J_{6,7}$	$J_{7,8}$	$J_{8,8a}$	
21α	S	10.7	4.9	9.3	10.3	9.3	4.9	
21β	S	11.2	5.4	10.8	9.3	9.3	9.3	
22	NH	11.2	5.4	10.7	8.5	8.9	8.7	

Attempts to prepare bicyclic azasugar analogues in which n = 2 (see compound 9) were not successful. Treatment of 17 with 1,4-butanediamine gave a mixture of four unstable products which were not analyzed further. On the other hand, 4-amino-1-butanol gave principally the monocyclic azasugar 25 whose structure was deduced by comparison of its 13 C NMR spectrum with that of the product 26 derived from reaction of N-methylamine with 17. Compound 25 had limited stability and was not characterized further.

In conclusion, bicyclic azasugar analogues of D-xylose containing a glycosidic heteroatom (N, O, and S) have been prepared and structurally characterized. Those compounds with a six-membered ring fused to the azasugar ring have long-term stability in aqueous solution, but those with fused five-membered rings, with the exception of the thiazolidine analogue, have limited stability. Those with fused seven-membered rings either do not form or are very unstable. When the glycosidic heteroatom is O or S, α -anomers are present in solution but the β -anomers predominate; when the heteroatom is N, only a β -anomer is present. α -Anomers favor having the glycosidic heteroatom axial to the azasugar ring while β -anomers have a very strong preference for the heteroatom to be equatorial. Results of the evaluation of the inhibitory potency and selectivity of these compounds will be reported separately.

EXPERIMENTAL

General

Reactions were run at room temperature unless indicated otherwise. Thin-layer chromatography (TLC) was run on Whatman Al Sil G/UV plates. Compounds were located on TLC plates by using molybdate spray reagent [Ce(SO₄)₂, 10 g; (NH₄)₆Mo₇O₂₄ 4H₂O, 25 g; H₂O, 900 mL; 98% H₂SO₄, 100 mL]. Flash column chromatography was performed on silica gel G (Fisher Scientific, S704-25, 60-200 mesh). Melting points were determined on a Thomas Hoover capillary melting point apparatus and are uncorrected. Elemental analyses were determined by M-H-W Laboratories in Phoenix, AZ. Fourier Transform Nuclear Magnetic Resonance (FT-NMR) spectra were recorded on a Varian 200, 300, or 500 MHz spectrometer; unless indicated otherwise, ¹H spectra were obtained at 500 MHz and ¹³C spectra at 125 MHz. HETCOR, COSY, and NOE experiments were used to make assignments of the ¹H and ¹³C NMR spectra. The NMR program 'gNMR' (Cherwell Scientific Publishing, Ltd., Oxford, U.K., 1996) was used to simulate the ¹H NMR peaks for the protons of the hydroxylated ring of compounds 18 \beta and 22 and H-2 through H-4 of 19, and the reported coupling constants and chemical shifts were obtained from these simulations. Mass spectra (MS) were determined using a Jeol JMS-SX102A double-focusing, high-resolution spectrometer; CH₅⁺ was used for positive ion chemical ionization (CI), and xenon was used as the FAB gas with thioglycerol as the matrix. High-resolution mass spectra (HRMS) were measured at 10,000 or better resolution and calibrated with highboiling perfluorokerosene (CI) or poly(ethylene glycol) (FAB). X-ray crystal and intensity data were collected using MoKa radiation ($\lambda = 0.71073\text{Å}$) and a Siemens R3m/V automated diffractometer for compound 19 and a Bruker P4 diffractometer for compound 20.

[7*R*-(7 α ,8 β ,9 α ,9a α)]-Octahydro-2*H*-pyrido[1,2-a]pyrimidine-7,8,9-triol (6). A solution of 17 (150 mg, 0.70 mmol) and 1,3-propanediamine (59 μ L, 0.70 mmol) in water (2 mL) was stirred for 10 min and then treated with Amberlite CG-400 ion exchange resin (hydroxide form) to remove HBr. Removal of the water under vacuum gave a slightly yellow syrup which gave colorless crystals of 6 (55 mg, 27%, mp 176.5-178°C) from ethanol. NMR spectral data for the product (6) were identical with those reported previously.¹⁰

5-Bromo-5-deoxy-1,2-*O*-isopropylidene-α-D-xylofuranose (16).¹⁵ A solution of 1,2-*O*-isopropylidene-5-*O*-tosyl-α-D-xylofuranose (14) (10.0 g, 29.1 mmol) and tetramethylammonium bromide (18.0 g, 117 mmol) in anhydrous DMF (180 mL) under a nitrogen atmosphere was heated at 120°C for 6 h. The reaction was followed by a TLC system of 40% EtOAc in hexane. A precipitate formed, and it was removed by filtration. The DMF was evaporated under vacuum, and the residue was purified by silica gel chromatography. Elution with 20% ethyl acetate in hexane gave 16 as a white powder (6.9 g, 94%, mp 99-100°C). ¹H NMR (200 MHz, CDCl₃, reference TMS) δ 5.96 (d, J = 3.4 Hz, 1H), 4.56 (d, J = 3.4 Hz, 1H), 4.43 (m, 2H), 3.51 (m, 2H), 1.93 (d, J = 5.2 Hz, OH, D₂O exchange), 1.52 (s, 3H), 1.33 (s, 3H); ¹³C NMR (CDCl₃, reference CDCl₃ δ 77.0) δ 112.1, 105.3, 85.0, 80.2, 74.4, 26.8, 26.8, 26.2. HRMS (CI): calcd for $C_8H_{13}BrO_4$: 253.0075 (M+1, ⁷⁹Br), found 253.0086; 255.0055 (M+1, ⁸¹Br), found 255.0056.

5-Bromo-5-deoxy- α -D-xylofuranose (17 α) and 5-bromo-5-deoxy- β -D-xylofuranose (17 β). A solution 16 (2.0 g, 7.9 mmol) in TFA (8 mL) and H₂O (2 mL) was stirred for 15 min. Water (15 mL) was added, and the solvent was removed under vacuum. Then another 15 mL water was added and evaporated. The same procedure was repeated two more times. The residue was dissolved in water and treated with Amberlite CG-400 ion exchange resin (hydroxide form) until the solution was neutralized. Removal of the water under vacuum gave 17 as a gum, which was a mixture of α - and β -anomers (1.6 g, 93%). The product crystallized from ethanol and chloroform (mp 79-81°C). At equilibrium in D₂O the product was an approximately 1:1 mixture of α - and β -anomers. 17 α ¹H NMR (D₂O, reference acetone δ 2.04) δ 5.29 (d, J =

4.4 Hz, 1H, H-1), 4.32 (m, 1H, H-4, overlaps with H-4 of β anomer), 4.14 (dd, J = 3.6, 5.2 Hz, 1H, H-3), 3.98 (dd, J = 3.6, 4.4 Hz, 1H, H-2), 3.41 (d, J = 10.7 Hz, 1H, H-5), 3.29 (dd, J = 7.3, 10.7 Hz, 1H, H-5); ¹³C NMR (75 MHz, D₂O, reference acetone δ 29.80) δ 95.7 (C-1), 78.1 (C-4), 75.4 (C-2), 74.8 (C-3), 29.8 (C-5). 17 β ¹H NMR (D₂O, reference acetone δ 2.04) δ 5.06 (br s, 1H, H-1), 4.30 (m, 1H, H-4, overlaps with H-4 of α anomer), 4.06 (dd, J = 2.1, 4.5 Hz, 1H, H-3), 3.96 (br s, 1H, H-2), 3.50 (dd, J = 5.8, 10.3 Hz, 1H, H-5), 3.40 (dd, J = 2.1, 10.3 Hz, 1H, H-5); ¹³C NMR (75 MHz, D₂O, reference acetone δ 29.80) δ 101.6 (C-1), 81.5 (C-4), 80.0 (C-2), 74.4 (C-3), 29.9 (C-5). Anal. Calcd for C₅H₉BrO₄: C, 28.19; H, 4.26. Found: C, 28.12; H, 4.43.

 $[7R-(7\alpha,8\beta,9\alpha,9a\beta)]$ -Hexahydro-2*H*,6*H*-pyrido[2,1-*b*][1,3]oxazine-7,8,9-triol (18 α) and [7*R*- $(7\alpha,8\beta,9\alpha,9a\alpha)$]-hexahydro-2*H*,6*H*-pyrido[2,1-*b*][1,3]oxazine-7,8,9-triol (18 β). A solution of 17 (100 mg, 0.47 mmol) and 3-amino-1-propanol (36 μ L, 0.47 mmol) in water (1 mL) was stirred for 10 min and then treated with Amberlite CG-400 ion exchange resin (hydroxide form) to remove HBr. Evaporation under vacuum gave a slightly yellow syrup which upon crystallization from ethanol gave colorless crystals that were only the β -anomer (31 mg, 35%, mp 141-141.5°C). At equilibrium in D₂O the ratio of α - (18 α) to β -anomers (18 β) was 12:88. 18 α ¹H NMR (D₂O, reference CH₃OH δ 3.30) δ 4.48 (d, J = 3.1, Hz, 1H, H-9a), 4.11 (dd, J= 4.8, 11.6 Hz, 1H, H-2eq), 3.75 (dt, J = 2.7, 11.6 Hz, 1H, H-2ax), 3.53²⁰ (obscured, 1H, H-7), 3.45 (t, J =9.8 Hz, 1H, H-8), 3.43^{20} (obscured, 1H, H-9), 3.13 (dt, J = 3.1, 13.7 Hz partly obscured, 1H, H-4ax), 3.09 (t, J = 11.0 Hz partly obscured, 1H, H-6ax), 2.92 (dm, J = 13.7 Hz, 1H, H-4eq), 2.64 (dd, J = 4.9, 11.0 Hz, 1H, H-6eq), 2.21 (m, 1H, H-3ax), 1.19 (br d, $J \sim 14.8$ Hz, 1H, H-3eq); ¹³C NMR (75 MHz, D₂O, reference CH₃OH δ 49.15) δ 89.0 (C-9a), 73.6 (C-8), 71.5 (C-9), 70.1 (C-7), 68.8 (C-2), 50.2 (C-4), 48.6 (C-6), 19.0 (C-3). 18 β ¹H NMR (D₂O, reference CH₃OH δ 3.30) δ 4.03 (dd, J = 4.8, 11.6 Hz, 1H, H-2eq), 3.55 (ddd, J = 4.9, 9.3, 10.7 Hz, 1H, H-7), 3.51 (m, obscured, 1H, H-2ax), 3.41 (d, J = 7.6, 9.4 Hz, 1H, H-9a), 3.28 (dd, J =9.3, 9.4 Hz, 1H, H-8), 3.25 (dd, J = 7.6, 9.4 Hz, 1H, H-9), 2.95 (br d, $J \sim 11.7$ Hz, 1H, H-4eq), 2.82 (dd, J = 7.6, 9.4 Hz, 1H, H-9), 2.95 (br d, $J \sim 11.7$ Hz, 1H, H-4eq), 2.82 (dd, J = 7.6, 9.4 Hz, 1H, H-9), 2.95 (br d, $J \sim 11.7$ Hz, 1H, H-4eq), 2.82 (dd, J = 7.6, 9.4 Hz, 1H, H-9), 2.95 (br d, $J \sim 11.7$ Hz, 1H, H-4eq), 2.82 (dd, J = 7.6, 9.4 Hz, 1H, H-9), 2.95 (br d, $J \sim 11.7$ Hz, 1H, H-4eq), 2.82 (dd, $J \sim 11.7$ Hz, 1H, H-4eq), 2.82 4.9, 11.7 Hz, 1H, H-6eq), 2.36 (ddd, J = 3.1, 12.0, 12.3 Hz, 1H, H-4ax), 2.13 (dd, J = 10.7, 11.7 Hz, 1H, H-6ax), 1.85 (m, 1H, H-3ax), 1.57 (br d, $J \sim 13.7$ Hz, 1H, H-3eq); ¹³C NMR (75 MHz, D₂O, reference CH₃OH δ 49.15) δ 93.8 (C-9a), 76.6 (C-8), 73.5 (C-9), 68.6 (C-7), 67.3 (C-2), 55.0 (C-6), 52.2 (C-4), 24.7 (C-3). Anal. Calcd for C₈H₁₅NO₄: C, 50.78; H, 7.99; N, 7.40. Found: C, 51.00: H, 7.78; N, 7.51.

[7*R*-(7α,8β,9α,9aα)]–Octahydro-1-methyl-2*H*-pyrido[1,2-*a*]pyrimidine-7,8,9-triol (19). A solution of 17 (107 mg, 0.50 mmol) and N-methyl-1,3-propanediamine (53 μL, 0.50 mmol) in water (2 mL) was stirred 10 min and then treated with Amberlite CG-400 ion exchange resin (hydroxide form) to remove HBr. ¹H NMR showed the presence of 19 and 20 in a ratio of 5:1. Evaporation of the water under vacuum gave a slightly yellow syrup part of which crystallized from 95% ethanol as a 1:7 mixture of 19 and 20 (6 mg). The filtrate was evaporated under vacuum and then crystallized from ethanol to give pure 19 (45 mg, 44%, mp 148.5-150.5°C). ¹H NMR (D₂O, reference CH₃OH δ 3.30) δ 3.46 (ddd, J = 4.9, 9.3, 10.7 Hz, 1H, H-7), 3.32 (dd, J = 8.8, 9.3 Hz, 1H, H-9), 3.26 (t, J = 9.3 Hz, 1H, H-8), 2.97 (d, J = 8.8 Hz, 1H, H-9a), 2.91 (ddt, J = 2.0, 3.9, 12.2 Hz, 1H, H-2eq), 2.88 (ddt, J = 2.0, 3.9, 13.2 Hz, 1H, H-4eq), 2.75 (dd, J = 4.9, 11.7 Hz, 1H, H-6eq), 2.75 (dt, J = 3.0, 13.2 Hz, 1H, H-4ax), 2.34 (s, 3H, CH₃), 2.24 (dt, J = 3.1, 12.2 Hz, 1H, H-2ax), 2.03 (dd, J = 10.7, 11.7 Hz, 1H, H-6ax), 1.94 (dddd, J = 3.9, 12.2, 13.2, 14.2 Hz, 1H, H-3ax), 1.31 (dddt, J = 2.0, 3.0, 3.1, 14.2 Hz, 1H, H-3eq); ¹³C NMR (75 MHz, D₂O, reference CH₃OH δ 49.15) δ 81.0 (C-9a), 77.6 (C-8), 70.4 (C-9), 68.4 (C-7), 57.5 (C-6), 54.9 (C-2), 52.5 (C-4), 33.0 (CH₃), 18.4 (C-3). Anal. Calcd for C₉H₁₈N₂O₃: C, 53.45; H, 8.97; N, 13.85. Found: C, 53.25; H, 8.85; N, 13.67.

[5S-(5α,7β,8α,9β,9aβ)]–Octahydro-7,8,9-trihydroxy-5-methyl-2*H*-pyrido[1,2-*a*]pyrimidin-5-ium bromide (20). A solution of 17 (126 mg, 0.59 mmol), triethylamine (0.25 mL, 1.8 mmol), and N-methyl-1,3-propanediamine (68 μL, 0.65 mmol) in water (2 mL) was stirred 10 min and then treated with Amberlite CG-400 ion exchange resin (hydroxide form) to remove HBr. ¹H NMR showed the presence of 19 and 20 in a ratio of 1:2. Evaporation of the water under vacuum gave a syrup which produced colorless crystals of 20 (70 mg, 41%, mp 209-210°C) from ethanol. ¹H NMR (D₂O, reference CH₃CN δ 1.94) δ 4.14 (d, J = 9.3 Hz, 1H, H-9a), 3.97 (ddd, J = 5.0, 9.3, 11.7 Hz, 1H, H-7), 3.52 (m, 1H, H-4eq), 3.49 (dd, J = 9.3, 9.3 Hz, 1H, H-9), 3.48 (dd, J = 5.0, 12.7 Hz, 1H, H-6eq), 3.45 (dd, J = 9.3, 9.3 Hz, 1H, H-8), 3.39 (dt, J = 3.4, 13.2, 13.2 Hz, 1H, H-4ax), 3.14 (dd, J = 5.1, 13.9 Hz, 1H, H-2ax), 3.05 (dd, J = 11.7, 12.7 Hz, 1H, H-6ax), 2.93 (s, 3H, CH₃), 2.77 (ddd, J = 3.9, 13.2, 13.7 Hz, 1H, H-2eq), 2.05 (m, 1H, H-3ax), 1.60 (br d, J = 15.6 Hz, 1H, H-3eq); ¹³C NMR (75 MHz, D₂O, reference CH₃CN δ 1.40) δ 85.6 (C-9a), 77.1 (C-8), 68.6 (C-9), 65.7 (C-4), 65.2 (C-7), 62.2 (C-6), 42.5 (C-2), 39.3 (CH₃), 19.9 (C-3). Anal. Calcd for C₉H₁₉N₂O₃Br: C, 38.18; H, 6.76; N, 9.89. Found: C, 38.14; H, 6.61; N, 9.66.

 $[6R-(6\alpha,7\beta,8\alpha,8a\beta)]$ -Hexahydro-5*H*-thiazolo[3,2-*a*]pyridine-6,7,8-triol (21α) [6R- $(6\alpha,7\beta,8\alpha,8a\alpha)$]-hexahydro-5H-thiazolo[3,2-a]pyridine-6,7,8-triol (21 β). A solution of 17 (398 mg, 1.88 mmol), 2-aminoethanethiol hydrochloride (218 mg, 1.92 mmol), and sodium carbonate (204 mg, 1.92 mmol) in water (4 mL) was stirred overnight and then evaporated under vacuum. The residue was chromatographed on silica gel eluting with 5% CH₃OH in CH₂Cl₂ first followed by 8% CH₃OH in CH₂Cl₂. The product was crystallized from ethanol as only the β-anomer (81 mg, 23%, mp 168-170°C). At equilibrium in D₂O solution, the product consisted of a 3:7 mixture of α - (21 α) and β -anomers (21 β). 21 α ¹H NMR (D₂O, reference $CH_3CN \delta 1.95) \delta 4.81 (d, J = 4.9 Hz, 1H, H-8a), 3.93 (dd, J = 4.9 9.3 Hz, 1H, H-8), 3.57 (dd, J = 9.3, 10.3)$ Hz, 1H, H-7), 3.48 (ddd, $J \sim 4.9$, 9.3, 10.7 Hz, 1H, H-6), 3.33 (m, 1H, H-3b), 2.98 (m, 1H, H-2), 2.90 (m, 1H, H-3a), 2.82 (m, 1H, H-2), 2.61 (dd, $J \sim 4.9$, 11.2 Hz, 1H, H-5eq), 2.45 (dd, $J \sim 10.7$, 11.2 Hz, 1H, H-5ax); ¹³C NMR (75 MHz, D_2O , reference $CH_3OH \delta 49.15$) $\delta 76.1$ (C-8a), 73.5 (C-7), 71.0 (C-8), 69.5 (C-6), 58.2 (C-3), 50.2 (C-5), 27.5 (C-2). **21** β ¹H NMR (D₂O, reference CH₃CN δ 1.95) δ 3.58 (ddd, J = 5.4, 9.3, 10.8 Hz, 1H, H-6), 3.46 (d, J = 9.3 Hz, 1H, H-8a), 3.29 (dd, J = 9.3, 9.3 Hz, 1H, H-8), 3.28 (m, 1H, H-3a), 3.21 (dd, J = 9.3) = 9.3, 9.3, 1H, H-7), 3.20 (dd, J = 5.4, 11.2 Hz, 1H, H-5eq), 2.91 (m, 2H, H-2), 2.52 (ddd, J ~ 7.6, 9.8, 9.8, 1H, H-3b), 2.17 (dd, $J \sim 10.8$, 11.2 Hz, 1H, H-5ax); ¹³C NMR (75 MHz, D₂O, reference CH₃OH δ 49.15) δ 78.3 (C-7), 75.4 (C-8), 71.6 (C-8a), 70.1 (C-6), 56.1 (C-3), 54.2 (C-5), 28.4 (C-2). Anal. Calcd for C₇H₁₃NO₃S: C, 43.96; H, 6.85; N, 7.32. Found: C, 43.83; H, 6.67; N, 7.06.

[6*R*-(6α,7β,8α,8aα)]–Octahydroimidazo[1,2-*a*]pyridine-6,7,8-triol (22). A solution of 17 and 1,2-diaminoethane in water was stirred for 0.5 h and then treated with CG-400 anion exchange resin (hydroxide form). The solvent was evaporated to give a gum whose 1 H and 13 C NMR spectra showed a major and a minor compound. The major was assigned structure 22: 1 H NMR (D₂O, reference CH₃OH δ 3.30) Major δ 3.63 (m, J = 5.4, 8.5, 10.7 Hz, 1H, H-6), 3.29 (dd, J = 8.5, 8.9 Hz, 1H, H-7), 3.26 (dd, J = 8.7, 8.9 Hz, 1H, H-8), 3.19 (dd, J = 5.4, 11.2 Hz, 1H, H-5eq), 3.06 (m, 1H, H-3b), 3.05 (m, 2H, H-2a and H-2b), 2.89 (d, J = 8.7 Hz, 1H, H-8a), 2.43 (dt, J ~ 8.3, 11.2, 1H, H-3a), 2.18 (dd, J = 10.7, 11.2 Hz, 1H, H-5ax); 13 C NMR (D₂O, reference CH₃OH δ 49.15) δ 80.2, 77.9, 73.6, 70.3, 52.4, 51.0, 43.2. Minor 13 C NMR (D₂O, reference CH₃OH δ 49.15) δ 82.4, 79.7, 75.4, 67.9, 62.8, 52.9, 39.3. HRMS (CI): calcd for C₇H₁₄N₂O₃: 175.1083 (M+1), found 175.1075.

Reaction of 5-bromo-5-deoxy-D-xylofuranose with 2-aminoethanol. A solution of 17 (30 mg, 0.14 mmol) and 2-aminoethanol (13 μ L, 0.21 mmol) in D₂O (0.7 mL) was monitored by ¹³C NMR which showed the formation of a major and a minor product in an approximate ratio of 2:1. Based on their ¹³C NMR chemical shifts, the major and minor appear to be 24 β and 24 α , respectively. ¹³C NMR (75 MHz, D₂O, reference CH₃OH δ 49.15) major: δ 94.3, 77.2, 73.1, 70.1, 66.4, 50.6, 49.9; minor: 92.4, 73.9, 73.5, 70.6, 69.4, 52.8, 51.6. HRMS (CI): calcd for C₇H₁₃NO₄: 176.0923 (M+1), found 176.0933.

Reaction of 5-bromo-5-deoxy-D-xylofuranose with 4-amino-1-butanol. A solution of 17 (27 mg, 0.13 mmol) and 4-amino-1-butanol (12 μ L, 0.13 mmol) in D₂O (0.7 mL) was monitored by ¹³C NMR which showed a single product that was assigned structure 25 based on comparison of its ¹³C NMR spectrum with that of compound 26. ¹³C NMR (75 MHz, D₂O, reference CH₃OH δ 49.15) δ 82.7, 73.9, 72.6, 70.0, 61.8, 52.4, 49.4, 29.7, 23.0. HRMS (CI): calcd for C₀H₁₉NO₅: 222.1341 (M+1), found 222.1343.

[2*R*-(2α,3α,4β,5α]-1-Methyl-2,3,4,5-piperidinetetrol (26). To a solution of 17 (25mg, 0.12mmol) in water (2 mL) was added 40% aqueous methylamine (30 μL, 0.36 mmol). After stirring for 10 min, the solvent was removed under vacuum. Then another 2 mL water was added and evaporated. The same procedure was repeated two more times to remove the excess methylamine. The residue was analyzed by NMR. ¹H NMR (D₂O, reference CH₃OH δ 3.30) δ 4.44 (d, J = 3.4 Hz, 1H, H-2), 3.42 (dd, J = 3.4, 9.3 Hz, 1H, H-3), 3.48 (dd, 1H, J = 9.3, 9.3 Hz H-4), 3.53 (ddd, J = 5.3, 9.3, 10.8 Hz, 1H, H-5), 2.63 (dd, J = 5.3, 11.2 Hz 1H, H-6eq), 2.54 (dd, J = 10.8, 11.2 Hz 1H, H-6ax), 2.31 (s, 3H, CH₃); ¹³C NMR (75 MHz, D₂O, reference CH₃OH δ 49.15) δ 84.5, 73.5, 72.4, 69.9, 50.8, 40.0. HRMS (FAB): calcd for C₆H₁₃NO₄: 164.0923 (M+1), found 164.0914.

X-ray Crystallographic Determinations The structures of 19 and 20 were solved using direct methods. It was apparent from the crystal data that the unit cell of 19 contained twelve molecules and in as much as the compound crystallized in the space group P2₁2₁2₁, that there are three molecules in the asymmetric unit. This was verified by the solution of 19. In the refinement process all nonhydrogen atoms of the two structures were refined anisotropically. Positions for all hydrogen atoms bonded to carbon atoms were calculated while positions for the hydrogens bonded to oxygen and nitrogen atoms were found in difference maps. All of the hydrogen atoms were allowed to ride on their neighboring heavy atom during the refinement process. The structure of 19 was solved using the program 'SHELXTL-PLUS' (Siemens Analytical X-ray Instruments, Inc., Madison, Wisconsin, 1990). The programs used in the refinement and display of 19 and the solution, refinement, and display of 20 are contained in the program package 'SHELXTL' PC, version 5.03 (Bruker Analytical X-ray, Madison, Wisconsin, 1994).

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- 15. The preparations of **15** and **17** have been described, but the latter product was not characterized. Tamura, T.; Kuneida, T.; Takizawa, T. J. Org. Chem. **1974**, 39, 38-44.
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- 17. The average bond lengths from the three molecules in the asymmetric unit are: N_1 - C_{9a} = 1.467Å and N_5 - C_{9a} = 1.480Å. For comparison, other average N-C bond lengths in this molecule are: N_1 - C_2 = 1.477Å, N_1 - C_{10} = 1.478Å, N_5 - C_4 = 1.474Å, and N_5 - C_6 = 1.458Å.
- 18. The bond lengths are: N₁-C_{9a} = 1.397Å and N₅-C_{9a} = 1.574Å. For comparison, other N-C bond lengths in this molecule are: N₁-C₂ = 1.466Å, N₅-C₄ = 1.530Å, N₅-C₅ = 1.493Å, and N₅-C₆ = 1.505Å.
 19. A similar transformation has been reported. The ¹³C NMR signals at 67.9 and 62.8 ppm distinguish the
- 19. A similar transformation has been reported.⁹ The ¹³C NMR signals at 67.9 and 62.8 ppm distinguish the minor compound from all the bicyclic azasugars we have prepared and characterized but correspond more closely with the 66.4 and 62.9 ppm reported for 1,4-dideoxy-1,4-imino-L-arabinitol, a monocyclic analogue of compound 23 reported in Nash, R. J.; Williams, J. M., Bell, E. A. *Phytochemistry* 1985, 24, 1620-1622.
- 20. Peak position obtained from cross peaks of COSY spectrum.